

Available online at www.sciencedirect.com



PHARMACOLOGY BIOCHEMISTRY ^{AND} BEHAVIOR

Pharmacology, Biochemistry and Behavior 75 (2003) 749-754

www.elsevier.com/locate/pharmbiochembeh

The effect of calcineurin activator, extracted from Chinese herbal medicine, on memory and immunity in mice

Jing Luo, Jiang-Hua Yin, Qun Wei*

Department of Biochemistry and Molecular Biology, Beijing Normal University, Beijing 100875, China Received 4 November 2002; received in revised form 5 February 2003; accepted 5 May 2003

Abstract

Calcineurin (CN) is a highly abundant phosphatase in the brain and it is the only Ca^{2+} and calmodulin-dependent protein serine/ threonine phosphatase. There is considerable evidence to suggest that CN plays an essential role in activity-dependent modulation of synaptic efficacy. It has been shown recently that inhibitors of CN, such as CsA or FK506, impair memory formation in day-old chicks. In our present study, extract of *Fructus cannabis* (EFC) with activation of CN, extracted from Chinese traditional medicine, was used to determine the effects on memory and immunity. In the step-down-type passive avoidance test, the plant extract (0.2 g/kg) significantly improved amnesia induced by chemical drugs in mice, and greatly enhanced the ability of cell-mediated type hypersensitivity and nonspecific immune responses in normal mice. The present study provided pharmacological evidence for Chinese herbal medicine screening from molecular model.

© 2003 Elsevier Inc. All rights reserved.

Keywords: Calcineurin; Activate; EFC; Step-down-type passive avoidance; Clearance of charcoal particles; Delayed-type hypersensitivity

1. Introduction

Protein phosphorylation and dephosphorylation are major mechanisms for controlling the activities of enzymes and other proteins (Rusnak and Mertz, 2000), and they are involved in regulating many life processes, such as memory formation and immune responses. The phosphorylation of kinases, such as the cAMP-dependent protein kinase A (PKA), the protein kinase C (PKC), the Ca²⁺/calmodulindependent protein kinase II (CaMK II) and the mitogenactivated protein kinase (MAPK), is critical for many forms of synaptic plasticity and for learning and memory (Abel et al., 1998; Kornhauser and Greenberg, 1997; Lee et al., 2000; Lisman, 1994). Although many studies have focused on the kinases in synaptic plasticity, relatively few reports have been conducted concerning phosphatases. But phosphorylation is a reversible process, and phosphatases are also essential enzymes to memory formation. Current neural network models of learning and memory formation also support that bidirectional modifications of synaptic plasticity is particularly essential to store information more effectively (Linden, 1994; Sejnowski, 1991).

Calcineurin (CN) is a highly abundant phosphatase in the brain and it is the only Ca²⁺ - and calmodulin-dependent protein serine/threonine phosphatase. CN has a narrower range of substrates than those of other phosphatases. CN dephosphorylates some important neuronal substrates, including cytoskeletal proteins such as tau (Wei et al., 2002; Kayyali et al., 1997), *N*-methyl-*D*-aspartate (NMDA) channel constituents (Lieberman and Mody, 1994) and the PKC substrates such as neurogranin and GAP-43 (neuromodulin) (Liu and Storm, 1989; Schrama et al., 1989; Seki et al., 1995). CN also dephosphorylates inhibitor 1 and DARPP32 (dopamine and cAMP-regulated phosphoprotein) (Hemmings and Greengard, 1986).

The studies on biological roles of CN have progressed to the important discovery that it is the common target of the immunosuppressant drugs cyclosporin A (CsA) and FK506 (Liu et al., 1991). They are pharmacological reagents that have been used to demonstrate CN as a major player in Ca^{2+} -dependent eukaryotic signal transduction pathways. During T cell activation, upon binding of antigen to T cell receptor (TCR), intracellular Ca^{2+} is elevated through the action of protein tyrosine kinases and phospholipase C. CN is activated through binding of Ca^{2+}/CaM and dephosphor-

^{*} Corresponding author. Tel.: +86-10-62208197(O); fax: +86-10-62207365.

E-mail address: weiq@bnu.edu.cn (Q. Wei).

vlate the cytosolic forms of the NF-ATc transcription factors. The dephosphorylated NF-ATc proteins translocate to the nucleus, where they bind, either alone or in cooperation with AP-1 family members, to specific cis elements in the promoter/enhancer regions of cytokine genes, such as IL-2. This pathway makes an important contribution to the induction of IL-2 gene transcription, a hallmark of T cell activation (Crabtree and Clipstone, 1994). CsA and FK506 inhibit CN activity after forming complexes with cytoplasmic immunophilins, cyclophilins and FKBP12, respectively (Kissinger et al., 1995; Mulkey et al., 1994; Clipstone et al., 1994; Trushin et al., 1994). These immunophilin-immunosuppressant complexes bind CN and inhibit its function by sterically hindering the access of substrates to the catalytic site (Luo et al., 1996; Griffith et al., 1995). The inhibition of CN is related to the adverse side effects of CsA and FK506.

In addition, CsA and FK506 are powerful probes in the research of CN's role in brain. Using CsA or FK506, most physiological studies including those conducted in our laboratories, show long-term potentiation (LTP) and longterm depression (LTD) are surveyed in hippocampal slices (Luo and Wei, 1998; Luo et al., 2002). But convincing pharmacobehavioural evidence for a role of CN in learning and memory formation is rather sparse. Up to now, there are few reports on single-trial, passive avoidance task in day-old chicks that have applied either CsA or FK506 as antagonists of CN. These reports indicate that the intracranial administration of CsA and FK506 disrupts memory formation. The effects were dose-dependent, most prominent when administered between 10 min before and 40 min after learning, and did not reach significance until at least 85 min posttraining whether 5 nm CsA was given immediately or 30 min posttraining, or 50 nm CsA was given immediately after training. Importantly, the memory impairment induced by CsA persisted for at least 24 h posttraining and was not apparent in chicks trained on 1 day with the drug administered 2 h prior to test on the second day. Hence, it is speculated that CsA selectively and permanently disrupts a relatively late stage of memory formation in the chick, possibly at least partially through inhibition of CN activity (Bennett et al., 1996; Bennett and Schmidt, 2002).

Except for some metal ions, such as Ca^{2+} , Mg^{2+} and Zn^{2+} , relatively little attention has been paid to the activator of CN. Based on the characters of CN, an effective molecular screening model was established in our laboratory (Yan and Wei, 1999). Using CN as a target enzyme, it is possible to determine its activity and its interaction with drugs. We then tested a number of traditional Chinese medicines (Yan et al., 2000), including *Fructus cannabis*.

F. cannabis is the seed of *Cannabis sativa* L. A number of major components have been identified in it, such as fat oil, lignanamide, campesterol stigmasterol, cannabinoid liganol, aminopeptidase, etc. It is a traditional remedy used to treat geriatric constipation. *F. cannabis* is reported to have a broad range of therapeutic effects including antitumor, antihypertensive and antihyperlipidaemic. In our laboratory, we

extracted a component of *F. cannabis* that activates CN. The effects of this extract of *F. cannabis* (EFC) on memory were investigated in one-trial passive avoidance step-down test in mice. Scopolamine, sodium nitrite and ethanol were chosen to inhibit memory. The muscarinolytic scopolamine only affects learning (Zhang and Saito, 1986). Sodium nitrite, which can induce anoxia in the brain, only affects retention (Kumar et al., 2000). The central nervous system depressant, 45% alcohol, disrupts the retrieval process (Vohra and Hui, 2000). The three chemicals can induce amnesia in step-down test, which is recommended as a simple, rapid and sensitive method in the study of learning and memory.

2. Materials and methods

2.1. Animals and housing conditions

Male Kunming mice, weighing 20 ± 2 g at the beginning of the experiment, were purchased from Experimental Animals Center of Beijing University. Animals were housed in group under the following laboratory conditions: temperature 20 ± 1 °C, humidity 40-60%, 12:12 L/D cycle, lights on at 08:00 h. Mice had free access to food and water, except during behavioral experiments. Animals were treated in accordance with the current law and the NIH Guide for Care and Use of Laboratory Animals.

2.2. Preparation of EFC

F. cannabis (500 g) was purchased from Beijing Tongrentang Pharmaceutical Group. Materials were extracted two times with 90% ethanol for 24 h and filtrated. The filtrate was concentrated to dry. The ethanol extract was subjected to column chromatography under reduced pressure on silica gel, eluted with a chloroform-methanol solvent system. The elutions that can activate CN were collected and dried (EFC), the total yield was 5 g (1%) in terms of starting materials.

Bovine brain CN and calmodulin were isolated from frozen tissues as previously described (Wei et al., 1993). CN activity was assayed using the method of our laboratory (Yan and Wei, 1999). The phosphatase activity of CN was defined as 100%. EFC activated the phosphatase activity of CN and induced a $35\pm5\%$ increase.

2.3. Drug administration

EFC was dissolved in saline prior to administration at doses of 0.2, 0.4 and 0.8 g/kg. Scopolamine hydrobromide (1.5 mg/kg) and sodium nitrite (NaNO₂, 120 mg/kg) were dissolved in sterile 0.9% saline prior to injection. The injection volume was kept constant at 10 ml/kg irrespective of dose; 45% ethanol was diluted in saline and orally administered in a volume of 20 ml/kg. Dinitrochlorobenzene (DNCB) was dissolved in acetone.

Table 1 Effects of EFC on transient amnesia of acquired learning induced by scopolamine

Treatment (g/kg) ^a	Step-down latency (s)	Number of errors
Saline	230.7±32.6**	$0.5 \pm 0.2 **$
Sco+saline	102.8 ± 30.5	1.4 ± 0.3
Sco + EFC (0.2)	$217.1 \pm 29.7*$	$0.6 \pm 0.2*$
Sco + EFC (0.4)	279.1±12.1***	0.4 ± 0.2 ***
Sco + EFC (0.8)	$257.0 \pm 24.3 ***$	$0.3 \pm 0.2^{***}$

All the values are expressed as mean \pm S.E.M., n = 11.

^a Mice were administrated saline or EFC (0.2, 0.4 and 0.8 g/kg ig) once daily for 7 days. Ten minutes before training on Day 6, saline or scopolamine was administered intraperitoneally in mice. The testing trial carried out on Day 7.

* P<.05.

** P<.01.

*** P<.001 vs. Sco+saline.

2.4. Step-down-type passive avoidance test

2.4.1. The effect of EFC on transient amnesia of acquired learning induced by scopolamine

After 7-day oral administration of EFC, long-term memory was examined using the step-down-type of passive avoidance test. The apparatus consisted of acrylic box with a stainless-steel grid floor. A platform was fixed in the end of the box. Electric shock (1 Hz, 1 ms, 36 V dc) was delivered to the grid floor with an isolated pulse stimulator. When mice were placed in the box, the electrical resistance varied between 100 and 250 k Ω . Therefore, each mouse received an electric shock varying between 0.14 and 0.36 mA. Ten minutes before training, saline or scopolamine was administered intraperitoneally in mice. At the beginning of training, mice were placed in the box to adapt for 3 min without electric shock. When electric shock was delivered, mice escaped from the grid floor back up onto the platform. The duration of training test was for 5 min and the shock was maintained for this period. Twenty-four hours after training, mice were placed on the platform for the retention test. The electric shocks were still delivered for 5 min. Step-down latency and the number of errors were recorded with improved retention reflected by increased latency and a reduction in errors.

2.4.2. The effect of EFC on transient amnesia of memory retention induced by sodium nitrite

After 7-day administration of EFC, saline or sodium nitrite was subcutaneously injected immediately after training. Twenty-four hours later, mice were placed on the platform for testing. Step-down latency and the number of errors were recorded within 5 min.

2.4.3. The effect of EFC on transient amnesia of memory retrieval induced by 45% ethanol

After 7-day administration of EFC, mice were trained. Thirty minutes before testing, saline or 45% ethanol was orally administered. Step-down latency and the number of errors were recorded within 5 min.

2.5. Clearance of charcoal particles

Mice were injected intravenously with 1:4 diluted Yidege ink 10 ml/kg. At 1 min (T_1) and 12 min (T_2), blood (20 µl) was taken from retro-ocular venous plexus and resolved in 0.1% Na₂CO₃ solution (2 ml). The absorbency (A) was measured at 675 nm in 720 spectrophotometer (Zhao et al., 1997). The clearance rate K and clearance index α were calculated:

$$K = (\lg C_1 - \lg C_2)/(T_1 - T_2) \quad \alpha = (W/WLS) \cdot K^{1/3}$$

where W = weight of rat and WLS = liver weight + spleen weight.

2.6. Delayed-type hypersensitivity

Kunming mice were depilated using BaS (barium sulfide) on the nape skin. 50% DNCB diluted in acetone was used to induce hypersensitivity. After 10-day administration of EFC, mice were attacked on the abdomen skin by 2.5% DNCB. Twenty-four hours later, mice were injected intravenously with 1% Evans Blue (10 ml/kg) and killed after 30 min. The blue-dyed skin on the abdomen was removed and macerated for 24 h in 4-ml solutions of acetone-physiological saline (1:1) mixture. Then the mixture was centrifuged and absorbency of the supernatants was measured at 610 nm in 720 spectrophotometer.

2.7. Statistical analysis

The data were expressed as mean \pm S.E.M. Statistical analysis of the data for multiple comparisons was performed by one-way ANOVA followed by the Dunnett's *t* test.

3. Results

3.1. Effects of EFC on transient amnesia of acquired learning induced by scopolamine

Effects of EFC on transient amnesia of acquired learning induced by scopolamine were shown in Table 1. Scopol-

Table 2

Effects of EFC on transient amnesia of memory retention induced by sodium nitrite

Treatment (g/kg) ^a	Step-down latency (s)	Number of errors
Saline	297.6±2.4**	$0.1 \pm 0.1*$
Sodium nitrite + saline	168.3 ± 44.1	0.9 ± 0.3
Sodium nitrite + EFC (0.2)	$283.4 \pm 14.5*$	$0.2 \pm 0.1*$
Sodium nitrite + EFC (0.4)	$278.6 \pm 14.4*$	0.3 ± 0.2
Sodium nitrite + EFC (0.8)	226.2 ± 33.4	0.4 ± 0.2

All the values are expressed as mean \pm S.E.M., n = 10.

^a Mice were administrated saline or EFC (0.2, 0.4 and 0.8 g/kg ig) once daily for 7 days. Saline or sodium nitrite was subcutaneously injected immediately after the training trial on Day 6. The testing trial carried out on Day 7.

* P<.05.

** P < .01 vs. sodium nitrite + saline.

Table 3 Effects of EFC on transient amnesia of memory retrieval induced by 45% ethanol

Treatment (g/kg) ^a	Step-down latency (s)	Number of errors
Saline	275.2±26.0***	$0.1 \pm 0.1 ***$
Ethanol + saline	49.5 ± 17.8	5.5 ± 1.3
Ethanol+EFC (0.2)	$162.8 \pm 42.0*$	$2.1 \pm 0.8*$
Ethanol $+$ EFC (0.4)	98.0 ± 38.3	3.8 ± 0.8
Ethanol + EFC (0.8)	120.9 ± 32.5	$1.6 \pm 0.5 **$

All the values are expressed as mean \pm S.E.M., n = 11.

^a Mice were administrated saline or EFC (0.2, 0.4 and 0.8 g/kg ig) once daily for 7 days. After 7-day administration of EFC, mice were trained. Thirty minutes before testing, saline or 45% ethanol was orally administered in mice.

** P<.01.

*** P <.001 vs. ethanol+saline.

amine (1.5 mg/kg) significantly shortened the latencies and increased the numbers of errors determined by step-down test. The extract (0.2, 0.4 and 0.8 g/kg) significantly improved dysmnesia mice performance.

3.2. Effects of EFC on transient amnesia of memory retention induced by sodium nitrite

Effects of EFC on transient amnesia of memory retention induced by sodium nitrite were shown in Table 2. Sodium nitrite impaired the step-down-type passive avoidance test performance of mice both in latencies and numbers of errors. EFC significantly prolonged the latencies at a dose of 0.2 and 0.4 g/kg, and decreased the numbers of error at a dose of 0.2 g/kg. The highest dose of extract (0.8 g/kg) also produced improvements but it was not statistically significant.

3.3. Effects of EFC on transient amnesia of memory retrieval induced by 45% ethanol

Effects of EFC on transient amnesia of memory retrieval induced by 45% ethanol were shown in Table 3; 45% ethanol impaired the step-down-type passive avoidance test performance of mice. EFC significantly prolonged the latency at a dose of 0.2 g/kg and decreased the number of errors at a dose of 0.2 and 0.8 g/kg. But a dose of 0.4 g/kg had no effects.

Table 4	
Effect of EFC on clearance of charcoal particles in normal mice	

Treatment (g/kg) ^a	Clearance rate (K)	Clearance index (α)
Saline	0.0245 ± 0.0021	4.5 ± 0.2
EFC (0.2)	$0.0315 \pm 0.0017 *$	$5.3 \pm 0.1 ***$
EFC (0.4)	0.0238 ± 0.0009	4.7 ± 0.1
EFC (0.8)	0.0247 ± 0.0022	4.7 ± 0.1

All the values are expressed as mean \pm S.E.M., n = 11.

 $^{\rm a}\,$ Mice were administrated saline or EFC (0.2, 0.4 and 0.8 g/kg ig) once daily for 7 days.

* P<.05.

*** P<.001 vs. saline.

 Table 5

 Effects of EFC on hypersensitive reaction in normal mice

Treatment (g/kg) ^a	Absorbency
Saline	0.155 ± 0.012
EFC (0.2)	0.209 ± 0.017
EFC (0.4)	0.202 ± 0.017
EFC (0.8)	$0.219 \pm 0.018*$

All the values are expressed as mean \pm S.E.M., n = 10.

 $^{\rm a}\,$ Mice were administrated saline or EFC (0.2, 0.4 and 0.8 g/kg ig) once daily for 10 days.

* P<.05, significant difference from saline treatment.

3.4. Effects of EFC on clearance of charcoal particles in normal mice

The effects of EFC on clearance of charcoal particles in normal mice were shown in Table 4. EFC (0.2 g/kg) produced significant increase on K and α .

3.5. The effects of EFC on hypersensitive reaction in normal mice

The effects of EFC on hypersensitive reaction in normal mice were shown in Table 5. EFC greatly increased the abilities of the hypersensitive reaction at a dose of 0.8 g/kg.

4. Discussion

In general, learning and memory are divided into three processes: learning acquisition, memory retention and retrieval. Chemical agents such as scopolamine, sodium nitrite and 45% ethanol impaired memory in mice trained on a step-down-type passive avoidance, which is used to measure the three stages of memory process depending on the drugtreated period. Scopolamine injected 10 min before training, sodium nitrite subcutaneously injected immediately after training trial and 45% ethanol orally administered 30 min before testing trial induced learning and memory impairment. The administration of EFC can improve the performance to some degree. EFC (0.2, 0.4 and 0.8 g/kg)produced an inverted bell-shaped dose effect on acquired learning of mice with scopolamine-induced dysmnesia. But the observed results of EFC at the same doses on other memory stages did not appear in the whole bell shape for retention and retrieval. Because EFC is an effective part of F. cannabis and contains a mixture of active components, EFC may be able to exert its improving effects through more than one component. It suggests that EFC might exert high effective action with an appropriate amount. Taking all these observations into account, including latencies and error numbers, we proposed that EFC can overcome amnesia in the three processes of learning and memory at a dose of 0.2 g/kg. Because CN potentially plays an essential role in activity-dependent modulation of synaptic efficacy, it is speculated that the improvement of EFC in memory is

^{*} *P* < .05.

753

related to activation of CN. It may be absorbed and activate CN. The activation of CN exerts subtle roles in brain via influencing LTP and LTD, which have been widely accepted as the synaptic models of learning and memory. LTP and LTD have been widely accepted as the synaptic models of learning and memory (Mulkey et al., 1994; Torii et al., 1995). In the CA1 region of the hippocampus, both LTP and LTD are induced by the increase of Ca^{2+} influx through the activated NMDA receptors (Tong et al., 1995). The highfrequency stimulation produces a great increase of Ca²⁺ influx, by which the PKC and Ca²⁺/CaMK II are activated and the induction of LTP is triggered. The low-frequency stimulation results in a moderate elevation of postsynaptic Ca^{2+} level, by which the CN is activated and LTD is induced. It has been suggested that CN may be activated more rapidly and in response to lower levels of Ca^{2+} than is CaMK II. At the same time, it has been argued that activitydependent alterations in intracellular Ca²⁺ concentrations may differentially affect the activity of CaMK II and CN, leading to changes in the phosphorylation state of key synaptic molecules, and hence to subtle and bidirectional changes in synaptic efficacy (Mansuy et al., 1998; Malleret et al., 2001; Winder et al., 1998; Lu et al., 1996, 2000; Zeng et al., 2001; Onuma et al., 1998). Furthermore, the activated CN may store memory more effectively. The CN-mediated effect of EFC on memory is just one possible explanation for the observed results. Other mechanism of action still cannot be excluded. The potential mechanism of EFC's improvement on memory may be a synergistic effect of active ingredients.

In our own laboratory, it had been found that immunosuppressive component from traditional Chinese drugs, ZIP1, could directly inhibit CN without drug-binding protein. At this stage, our experiment supports that EFC with activation of CN could increase the ability of immunity.

In sum, the administration of EFC significantly improves memory deficits in amnesic mice induced by chemical substance and increases the ability of immunity. At this stage, any conclusion that the improving effects of EFC are due to activation of CN must remain somewhat speculative. It is thus imperative to investigate the changes of activity and content of CN in brain. The precise mechanisms of action by which EFC improves memory and immunity also need further investigation but the present study provided direct pharmacobehavioural evidence for regulators screening from Chinese herbal medicine. More importance is that CN is an efficient enzyme for screening. The natural products we obtained from our screening offer the clues for further development of lead compounds.

Acknowledgements

This work was supported by the National Natural Science Foundation of China and the State Key Basic Research and Development Program.

References

- Abel T, Martin KC, Bartsch D, Kandel ER. Memory suppressor genes: inhibitory constraints on the storage of long-term memory. Science 1998;279:338–41.
- Bennett PC, Schmidt L. Cylosporin A, FK506 and rapamycin produce multiple, temporally distinct effects on memory following single-trial, passive avoidance training in the chick. Brain Res 2002;927:180–94.
- Bennett PC, Zhao WQ, Lawen A, Ng KT. Cylosporin A, an inhibitor of calcineurin, impairs memory formation in day-old chicks. Brain Res 1996;73:107–17.
- Clipstone NA, Fiorentino DF, Crabtree GR. Molecular analysis of the interaction of calcineurin with drug–immunophilin complexes. J Biol Chem 1994;269:26431–7.
- Crabtree GR, Clipstone NA. Signal transmission between the plasma membrane and nucleus of T lymphocytes. Annu Rev Biochem 1994;63: 1045-83.
- Griffith JP, Kim JL, Kim EE, Sintchak MD, Thomson JA, Fitzgibbon MJ, et al. X-ray structure of calcineurin inhibited by the immunophilin– immunosuppressant FKBP12–FK506 complex. Cell 1995;82:507–22.
- Hemmings Jr HC, Greengard P. DARPP-32, a dopamine-regulated phosphoprotein. Prog Brain Res 1986;69:149–59.
- Kayyali US, Zhang W, Yee AG, Seidman JG, Potter H. Cytoskeletal changes in the brains of mice lacking calcineurin Aα. J Neurochem 1997;68:1668–77.
- Kissinger CR, Parge HE, Knighton DR, Lewis CT, Pelletier LA, Tempczyk A, et al. Crystal structure of human calcineurin and the human FKBP12-FK506-calcineurin complex. Nature 1995;378:641-4.
- Kornhauser JM, Greenberg ME. A kinase to remember: dual roles for MAP kinase in long-term memory. Neuron 1997;18:839–42.
- Kumar V, Singh PN, Muruganandamav, Bhattacharya SK. Effect of Indian *Hypericum perforatum* Linn on animal models of cognitive dysfunction. J Ethnopharmacol 2000;72:119–28.
- Lee HK, Barbarosie M, Kameyama K, Bear MF, Huganir RL. Regulation of distinct AMPA receptor phosphorylation sites during bidirectional synaptic plasticity. Nature 2000;405:955–9.
- Lieberman DN, Mody I. Regulation of NMDA channel function by endogenous Ca²⁺-dependent phosphatase. Nature 1994;369:235–9.
- Linden DJ. Long-term synaptic depression in the mammalian brain. Neuron 1994;12:457–72.
- Lisman J. The CaM kinase II hypothesis for the storage of synaptic memory. Trends Neurosci 1994;17:406–12.
- Liu YC, Storm DR. Dephosphorylation of neuromodulin by calcineurin. J Biol Chem 1989;264:12800–4.
- Liu J, Farmer Jr JD, Lane WS, Friedman J, Weissman I, Schreiber SL. Calcineurin is a common target of cyclophilin–cyclosporin A and FKBP–FK506 complexes. Cell 1991;66:807–15.
- Lu YF, Tomizawa K, Moriwaki A, Hayashi Y, Tokuda M, Itano T, et al. Calcineurin inhibitors, FK506 and cyclosporin A, suppress the NMDA receptor-mediated potentials and LTP, but not depotentiation in the rat hippocampus. Brain Res 1996;729:142–6.
- Lu YM, Mansuy IM, Kandel ER, Roder J. Calcineurin-mediated LTD of GABAergic inhibition underlies the increased excitability of CA1 neurons associated with LTP. Neuron 2000;26:197–205.
- Luo J, Wei Q. Relationship between LTP and nuclear protein induced by calcineurin. Chin Sci Bull 1998;43:1384–7.
- Luo C, Shaw KT, Raghavan A, Aramburu J, Garcia-Cozar F, Perrino BA, et al. Interaction of calcineurin with a domain of the transcription factor NFAT1 that controls nuclear import. PNAS 1996;93:8907-12.
- Luo J, Lou AL, Wei Q. Effects of calcineurin on LTP of rats in vivo. Chin Sci Bull 2002;47:476–9.
- Malleret G, Haditsch U, Genoux D, Jones MW, Bliss TVP, Vanhoose AM, et al. Inducible and reversible enhancement of learning, memory and long-term potentiation by genetic inhibition of calcineurin. Cell 2001; 104:675–86.
- Mansuy IM, Mayford M, Jacob B, Kandel ER, Bach ME. Restricted and regulated overexpression reveals calcineurin as a key component in

the transition from short-term to long-term memory. Cell 1998;92: 39-49.

- Mulkey RM, Endo S, Shenolikar S, Malenka RC. Involvement of a calcineurin/inhibitor-1 phosphatase cascade in hippocampal long-term depression. Lett Nature 1994;369:486–8.
- Onuma H, Lu YF, Tomizawa K, Moriwaki A, Tokuda M, Hatase O, et al. A calcineurin inhibitor, FK506, blocks voltage-gated calcium channel-dependent LTP in the hippocampus. Neurosci Res 1998;30:313–9.
- Rusnak F, Mertz P. Calcineurin: form and function. Physiol Rev 2000;80: 1483-521.
- Schrama LH, Heemskerk FMJ, de Graan PNE. Dephosphorylation of protein kinase C phosphorylated B-50/GAP-43 by the calmodulin-dependent phosphatase calcineurin. Neurosci Res Commun 1989;5:141–7.
- Sejnowski TJ. Synaptic mechanisms for long-term depression. Curr Biol 1991;1:38-40.
- Seki K, Chen HC, Huang KP. Dephosphorylation of protein kinase C substrates, neurogranin, neuromodulin and MARCKS, by calcineurin and protein phosphatases 1 and 2A. Arch Biochem Biophys 1995;316: 673–9.
- Tong G, Shepherd D, Jahrt CE. Synaptic desensitization of NMDA receptors by calcineurin. Science 1995;267:1510–2.
- Torii N, Kamishita T, Otsu Y, Tsumoto T. An inhibitor for calcineurin, FK506, blocks induction of long-term depression in rat visual cortex. Neurosci Lett 1995;185:1–4.
- Trushin SA, Pennington KN, Algeciras-Schimnich A, Paya CV. Protein kinase C and calcineurin synergize to activate IκB kinase and NF-κB in T lymphocytes. J Biol Chem 1994;274:22923–31.

- Vohra BP, Hui X. Improvement of impaired memory in mice by taurine. Neural Plast 2000;7:245–59.
- Wei Q, Lu ZJ, Xiao FX, Xing LZ, Li DH. A useful method of calcineurin, calmodulin, and other calcium binding proteins. Chin Biochem J 1993; 9:6–10.
- Wei Q, Holzer M, Brueckner MK, Liu Y, Arendt T. Dephosphorylation of tau protein by calcineurin (PP2B) led to neural living cells. Cell Mol Neurobiol 2002;22:13–24.
- Winder DG, Mansuy IM, Osman M, Moallem TM, Kandel ER. Genetic and pharmacological evidence for a novel intermediate phase of long-term potentiation suppressed by calcineurin. Cell 1998;92:25–37.
- Yan LJ, Wei Q. High activity of the calcineurin A subunit with a V314 deletion. Biol Chem 1999;380:1281-5.
- Yan LJ, Yu CJ, Wei Q. Effect of different immunosuppressive drugs on calcineurin and its mutants. Sci China C 2000;43:68–74.
- Zeng HK, Chattarji S, Barbarosie M, Rondi-Reig L, Philpot BD, Miyakawa T, et al. Forebrain-specific calcineurin knockout selectively impairs bidirectional synaptic plasticity and working/episodic-like memory. Cell 2001;107:617–29.
- Zhang JT, Saito H. Studies on susceptibilities to the amnestic effects of 12 chemicals on passive avoidance responses in mice: comparison between step down and step through tests. Acta Pharm Sinica 1986;21:12–9.
- Zhao HW, Li YH, Zhu YN, Weng SA, Zhang Y, Ge SW. Effect of brucine on mouse nonspecific immune responses. Acta Pharmacol Sinica 1997; 18:468–70.